

DIVERSITY OF BACTERIA AND FUNGI IN THE PREPUCE OF CAMELS (*Camelus dromedarius*)

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ABSTRACT

A total of 42 infertile male dromedary camels (4-12 years old, 500-800 kg) and 7 young male camels (control, 2-2.5 years old) were enrolled in the present study. All camels were sedated and preputial swabs were collected and immediately immersed in 1 ml sterile solution of 0.9% NaCl. Preputial swabs were transferred refrigerated within 1 hour to the bacteriological laboratory. After preputial swabbing, semen samples were collected from infertile camels using an electro-ejaculator and evaluated for routine semen quality parameters using the conventional methods. Statistical analyses were conducted by Student's *t*-test and Chi-square (χ^2). Results revealed that semen parameters were reduced in infertile camels with preputial contamination. The bacterial count in 1 ml swab sample was 148×10^3 and 0.24×10^3 in infertile mature camels and control immature camels, respectively. Nine bacterial species isolated from 49 camels' preputial swabs comprised 41 gram-positive species (83.7%) and 8 gram-negative species (16.3%). The swabs of infertile camels had colonies species of *Staphylococcus*, *Streptococcus*, *Corynebacterium*, *Bacillus*, *E. coli*, *Pseudomonas aeruginosa*, *Actinomyces*, *Fusobacterium*, *Clostridium perfringens* at percentages of 35.72% (n=15), 21.43% (n=9), 11.91% (n=5), 9.52% (n=4), 7.14% (n=3), 4.76% (n=2), 4.76% (n=2), 2.38% (n=1), 2.38% (n=1), respectively. *Candida* colonies appears in concurrence with bacterial colonies in 28.57% (n=12). In control camels, the examined samples had colonies of *Streptococcus* spp., *Bacillus* spp. and *E. coli* spp. at proportions of 42.86% (n=3), 42.86% (n=3), 14.28% (n=1), respectively. *Candida* spp. observed in 5 colonies (71.43%) of control camels. In conclusion, *Staphylococcus*, *Streptococcus* and *Bacillus* were the frequently isolated bacterial spp. from prepuce of camel.

Key words: Bacterial diversity, camels, fungus, prepuce

The reproductive efficiency of camels under natural conditions is generally considered as low (Al-Qarawi, 2005) and genital infection is considered as most important reason (Tibary *et al*, 2006). It has been demonstrated that the preputial sac can act as a reservoir of organisms and it thus responsible for causing ascending uro-genital infection (Agartan *et al*, 2005). The preputial microbial community plays a key role in maintaining health and altered microbial communities have been associated with a variety of reproductive diseases (Nelson *et al*, 2010; Sandal and Inzana, 2010; Chaban *et al*, 2012). The source of camel preputial contamination is usually soil, faeces and female genital tract (Wickware *et al*, 2020), and microorganisms gain entry through preputial orifice (Paray *et al*, 2018). The camel often does not show any clinical signs of contamination,

but infection is traced back through symptoms shown in female camels it has mated (Al-Qarawi, 2005). A long duration of semen collection and collection in a sitting position (El-wishy, 1988) increase the risk of contamination of prepuce of camel. Bacterial contamination leads to a series of alterations including reduced sperm motility, morphology, various semen quality parameters (Najee *et al*, 2012; Perumal *et al*, 2013), and subsequent reduced fertility (Ochsendrof and Fuchs, 1993; Griveau *et al*, 1995). This study investigates the frequency of isolated bacteria and fungi in camels' prepuce.

Materials and Methods

Experimental animals

A total of 42 infertile male dromedary camels (4-12 years old, 500-800 kg) admitted to the

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Veterinary Teaching Hospital, King Faisal University and 7 young male camels (control, 2-2.5 years old) belonging to the Camel Research Centre, King Faisal University (25°23'N 49°36'E) were selected in the present study. The control males had no contact with females before or during the experiment (5 months) between November 2020 to and March 2021.

Samples collection

Infertile camels were restrained in sternal position with ropes, sedated intravenously with a combination of xylazine (Rompun®, Bayer; 0.15 mg/kg) and ketamine (Alfasan® 10%, Holland; 2.5 mg/kg) (White *et al*, 1982). Then, sedated camels were turned to lateral recumbency and their prepuce was cleaned externally with sterile saline solution (0.9% NaCl). A sterile swab was inserted and rotated in the prepuce, and immediately immersed in a sterile 5 ml plastic tubes containing 1 ml sterile physiological saline solution (0.9% NaCl). Preputial swabs were collected from the control males as in the infertile camels. All collected swabs were transferred refrigerated within 1 hour to the bacteriological laboratory in the Ministry of Agriculture, Al-Ahsa, Kingdom of Saudi Arabia. After preputial swabbing, semen samples were collected from all infertile camels using an electro-ejaculator (Tingari *et al*, 1986). Semen samples were evaluated for routine semen quality parameters such as volume (mL), viscosity (Ghoneim *et al*, 2010), pH, per cent motile sperm, sperm concentration ($\times 10^6$ /ml), and per cent sperm abnormalities (Table 1) by the same trained researcher using Sperm Vision 3.5 (Minitube of America, Inc.).

Table 1. Semen parameters of infertile mature camels (mean \pm SEM; range).

Semen parameters	Infertile mature camels (n=42) (Range)	Reference*
Volume (ml)	6.32 \pm 0.74 (7 - 25)	6.56 \pm 1.73
Viscosity (0 - 5)	3.57 \pm 0.25 (0 - 5)	4.25 \pm 0.37
pH	8.11 \pm 0.06 (6.8 - 8.4)	7.84 \pm 0.14
Motility (%)	8.00 \pm 2.11 (0 - 20)	43.13 \pm 9.77
Sperm concentration ($\times 10^6$ /ml)	37.07 \pm 13.41 (1 - 100)	366.15 \pm 47.55
Sperm abnormalities (%)	50.54 \pm 4.65 (15 - 80)	26.00 \pm 2.95

*Adapted from Waheed *et al* (2015).

Microbiological evaluation

Bacterial identification

Each swab sample was plated on 5% sheep blood agar, Mac-Conkey agar and *Salmonella-Shigella* agar and incubated at 37°C in 5% CO₂ for 24 h. Growing colonies were examined with Gram staining methods. Suspected colonies were identified with biochemical and carbohydrate fermentation tests using bioMérieux's API identification products (Chesbrough, 2004).

Counting colony forming units per millilitre (CFU/ml) of sample

The quantitative estimation of the microbial contamination of prepuce was performed by the standard plate count method (Shukla, 2011). An aliquot (0.05 ml) of the swab sample was diluted with 1:10 PBS (0.1M phosphate buffer containing 0.15M NaCl, pH 7.3), producing a serial-dilution. Plates of Columbia-blood-agar were inoculated with 100 μ l of the diluted swab sample (10¹-10⁵) and incubated at 37°C for 48 h. The bacterial colonies were counted with the help of a colony counter. The colonies were counted in relation to the swab sample aliquot of 1ml and expressed as CFU/ml.

Mycological examinations

Swab samples were examined for fungus and yeast by seeding the samples in Sabaroud's dextrose agar. The plates were incubated at 25°C for 5 days and checked for fungal growth from the fourth day onward. Fungal identification relied on the morphologic and physiologic features (de Hoog *et al*, 2000). Yeasts were sub-cultured to obtain pure cultures for identification. For definitive identification of yeasts, the carbohydrate assimilation pattern was defined by ID32 (Biomerieux, Bagno a Ripoli, Italy).

Statistical analysis

The data are presented as the means \pm SEM and percentages. Analyses were conducted by Student's *t*-test and Chi-square (χ^2) using INSTAT software 3.1 (2017).

Results and Discussion

The bacterial count in 1 ml swab sample was 148 $\times 10^3$ and 0.24 $\times 10^3$ in infertile mature camels and control immature camels, respectively (Table 2). Nine bacterial species isolated from 49 camels' preputial swabs comprised 41 gram-positive species (83.7%) and 8 gram-negative species (16.3%). The different types of colonies were *Staphylococcus* spp., *Streptococcus* spp., *Corynebacterium* spp., *Bacillus*

spp., *E. coli* spp., *Pseudomonas aeruginosa* spp., *Actinomyces* spp., *Fusobacterium* spp., *Clostridium perfringens* spp. and *Candida* spp. as shown in table 2. The swabs of infertile camels had colonies species of *Staphylococcus*, *Streptococcus*, *Corynebacterium*, *Bacillus*, *E. coli*, *Pseudomonas aeruginosa*, *Actinomyces*, *Fusobacterium*, *Clostridium perfringens* at percentages of 35.72% (n=15), 21.43% (n=9), 11.91% (n=5), 9.52% (n=4), 7.14% (n=3), 4.76% (n=2), 4.76% (n=2), 2.38% (n=1), 2.38% (n=1), respectively. *Candida* colonies appear in concurrent with bacterial colonies in 28.57% (n=12). In control camels, the examined samples had colonies of *Streptococcus* spp., *Bacillus* spp. and *E. coli* spp. at proportions of 42.86% (n=3), 42.86% (n=3), 14.28% (n=1), respectively. *Candida* spp. observed in 5 colonies (71.43%) of control camels (Table 2).

Table 2. Bacterial load, types and frequency in dromedary preputial samples.

Isolates	Infertile mature camels (N=42)	Control immature camels (N=7)
CFU/ml (mean \pm SEM)	148 ^a \pm 56.96x10 ³	0.24 ^b \pm 0.12x10 ³
<i>Staphylococcus</i> spp. (%)	11 (26.19%)	—
<i>Staphylococcus</i> spp. + <i>Candida</i> (%)	4 (9.52%)	—
<i>Streptococcus</i> spp. (%)	5 (11.91%)	—
<i>Streptococcus</i> spp. + <i>Candida</i> (%)	4 (9.52 ^a %)	3 (42.86 ^b %)
<i>Corynebacterium</i> spp. (%)	5 (11.91%)	—
<i>Bacillus</i> spp. (%)	3 (7.14 ^a %)	2 (28.57 ^b %)
<i>Bacillus</i> spp. + <i>Candida</i> (%)	1 (2.38 ^a %)	1 (14.28 ^b %)
<i>E. coli</i> spp. (%)	3 (7.14%)	—
<i>E. coli</i> spp. (%) + <i>Candida</i> (%)	—	1 (14.28%)
<i>Pseudomonas aeruginosa</i> (%)	2 (4.76%)	—
<i>Actinomyces</i> spp. + <i>Candida</i> (%)	2 (4.76%)	—
<i>Fusobacterium</i> + <i>Candida</i> (%)	1 (2.38%)	—
<i>Clostridium perfringens</i> (%)	1 (2.38%)	—

Means and percentages with dissimilar superscripts in the same row are significantly different at P<0.05.

In the present study, semen parameters were reduced in infertile camels. These reduced values were accompanied with preputial contamination in these camels. This relationship indicates that preputial microbial load might be considered as a relevant cause of infertility in dromedaries. Previous studies achieved the same findings (Ochsendrof and Fuchs, 1993; Griveau *et al*, 1995; Najee *et al*, 2012; Perumal *et al*, 2013). Published data on the presence and distribution of bacteria and fungi in

the camel prepuce are scarce. The current study revealed that the bacterial count in 1 ml semen was 0.24 and 148 $\times 10^3$ CFU in control immature and infertile camels, respectively. The young bulls had significantly lower total bacterial counts than older animals (Reddy *et al*, 1971). However, Brown *et al* (1974) found no correlation between the age and total bacterial count. The saprophytic flora of the prepuce of healthy bulls includes numerous bacterial species (Thibier and Guerin, 2000). The collected preputial samples were colonised by species of *Staphylococcus*, *Streptococcus*, *Corynebacterium*, *Bacillus*, *E. coli*, *Pseudomonas*, *Actinomyces*, *Fusobacterium* and *Clostridium*. Species of *Staphylococcus*, *Streptococcus*, *Corynebacterium* and *Bacillus* were the frequently isolated bacteria. Twelve bacterial spp. were identified in dromedary camels' prepuce that were *Serratia liquefaciens*, *Staphylococcus aureus*, *Streptococcus* spp., *Klebsiella ozaenae*, *Pseudomonas* spp., *Shigella* spp., *Enterobacter cloaceae*, *Flavobacterium* spp., *Actinomyces* spp., *Acinetobacter* spp., *Acinetobacter calcoaceticus* and *Bacillus* spp. (Serin *et al*, 2010). *Streptococcus* spp., *Pseudomonas* spp. and *Staphylococcus* spp. are the most common bacteria isolated from the uterus of infertile camels (Wernery and Wernery, 1992; Ali *et al*, 2010; Almohasen, 2011). Then, camel preputial contamination might lead to uterine infection and *vice versa*. It has been demonstrated that the camel prepuce can act as a reservoir of organisms and it thus might be responsible for infertility (Agartan *et al*, 2005). *Staphylococcus aureus* was the most frequently isolated aerobic microorganism in specimens obtained from the prepuce of dogs (Ling and Ruby, 1978). The present study was done during the camels' rutting season with a marked peak in sexual activity. This activity might lead to preputial contamination (Zhao, 2000; Fatnassi *et al*, 2014). In the present study, *Candida* spp. was colonised in 26.5% of the collected samples. *Candida* is a common mycoflora present in the genitalia of healthy female camels (Shokri *et al*, 2010). In camels mating, the penis penetrates the cervical canal, and in some cases, it enters deep into the uterine cavity (Serin *et al*, 2010). During the camels' rutting season, the atmospheric concentration of fungus spores is twice as high as in the non-rutting season (Bartzokas, 1975). In stallions, yeasts were isolated in 9.1% of the preputial samples (Rota *et al*, 2011). In conclusion, species of *Staphylococcus*, *Streptococcus* and *Bacillus* were the frequently isolated bacteria from camels' prepuce. The present study recommends the washing of the camel's prepuce before mating.

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References

Agartan CA, Kaya DA, Ozturk CE and Glucan A. Is aerobic preputial flora age dependent? *Japanese Journal of Infectious Diseases*. (2005); 58:276-278.

Ali A, Hassanein KM, Al-sobayil, FA, Tharwat M, Al-Hawas A and Ahmed AF. Relationship between characters of vaginal discharges and uterine bacterial isolates recovered from repeat breeding female camels (*Camelus dromedarius*). *Journal of Agriculture and Veterinary Sciences*, Qassim University. (2010) 2:87-97.

Almohasen FM. Uterine histopathological findings of infertile female camels (*Camelus dromedarius*) admitted to the Veterinary Teaching Hospital. Master Degree of Veterinary Sciences (M.V.Sc.) in Theriogenology, King Faisal University, Saudi Arabia. (2011).

Al-Qarawi AA. Infertility in the dromedary bull: A review of causes, relations and implications. *Animal Reproduction Science*. (2005); 87:73-92.

Bartzokas C. Relationship between the metereological conditions and the air-borne fungal flora of the Athens metropolitan area. *Mycopathologia*. (1975); 57:35-38.

Brown VG, Schollum LM and Jarvis BDW. Microbiology of bovine semen and artificial breeding practices under New Zealand conditions. *New Zealand Journal of Agriculture Research*. (1974); 17:431-442.

Chaban B, Chu S, Hendrick S, Waldner C and Hill JE. Evaluation of a *Campylobacter foetus* subspecies *venerealis* real-time quantitative polymerase chain reaction for direct analysis of bovine preputial samples. *Canadian Journal of Veterinary Research*. (2012); 76(3):166-173.

Chesbrough M. District laboratory practice in tropical countries, Cambridge University press, UK. (2004); pp 435.

El-wishy AB. Reproduction in the male dromedary (*Camelus dromedarius*): A review. *Animal Reproduction Science*. (1988); 17:217-241.

Fatnassi M, Padalino B, Monaco D, Khorchani T, Lacalandra GM and Hammadi M. Evaluation of sexual behaviour of housed male camels (*Camelus dromedarius*) through female parades: correlation with climatic parameters. *Tropical Animal Health and Production*. (2014); 46:313-321.

Ghoneim I, Al-Eknah M, Waheed MM, Alhaider AK and Homeida AM. Effect of some extenders and enzymes on semen viscosity and sperm viability in dromedary camels. *Journal of Camel Practice and Research*. (2010); 17:85-89.

Griveau JF, Domount E, Renard P, Challegani JP and Lelannou D. Reactive oxygen species lipid peroxidation and enzymatic defense system in human spermatozoa. *Journal of Reproduction and Fertility*. (1995); 103(1):17-26.

de Hoog GS, Guarro J, Gené J and Figueras MJ. *Atlas of Clinical Fungi*, 2nd Edition. Utrecht, Centraalbureau voor Schimmelcultures. (2000).

INSTAT. *INSTAT 3.1 General Statistical Package Software*. (2017).

Ling GV and Ruby AL. Aerobic bacterial flora of the prepuce, urethra and vagina of normal adult dogs. *American Journal of Veterinary Research*. (1978); 39(4):695-698.

Najee HB, Al-Shawii AM and Abd-Al Rahman LY. Bacterial contamination of imported bulls' frozen semen. *Al-Anbar Journal of Veterinary Science*. (2012); 5(1):1999-6527.

Nelson DE, Van Der Pol B, Dong Q, Revanna KV, Fan B, Easwaran S, Sodergren E, Weinstock GM, Diao L and Fortenberry JD. Characteristic male urine microbiomes associate with asymptomatic sexually transmitted infection. *PLoS One*. (2010); 5(11):e14116.

Ochsendorf FR and Fuchs J. Oxidative imbalances in male fertility. In: Fuchs J, Packer L, editors. *Oxidative stress in dermatology*. New York: Marcel Dekker. (1993); pp 489-531.

Paray AR, Bhakat M, Lone SA, Mohanty RS, Rahman JU, Khanday ZB and Danish Z. Role of preputial washing in reducing microbial load and improving bovine semen quality. *Asian Pacific Journal of Reproduction*. (2018); 7(3):97-102.

Perumal P, Kumar TK and Srivastava SK. Infectious causes of infertility in buffalo bull (*Bubalus bubalis*). *Buffalo Bulletin*. (2013); 32(2):71-96.

Reddy BT, Krishnamurthy DS and Venkataswami V. Bacterial flora of prepuce and the effect of intra-preputial treatment on the bacteriological quality of semen. *Indian Veterinary Journal*. (1971); 48:722-727.

Rota A, Calicchio E, Nardoni S, Fratini F, Ebani VV, Sgorbini M, Panzani D, Camillo F and Mancianti F. Presence and distribution of fungi and bacteria in the reproductive tract of healthy stallions. *Theriogenology*. (2011); 76:464-470.

Sandal I and Inzana TJ. A genomic window into the virulence of *Histophilus somni*. *Trends in Microbiology*. (2010); 18:90-99.

Serin I, Ceylan A, Kirkan S and Parin U. Preputial bacterial flora and antibiotic susceptibility in wrestling dromedary bulls in Aydin region of Turkey. *Journal of Animal and Veterinary Advances*. (2010); 9:482-485.

Shokri H, Khosravi A, Sharifzadeh A and Tootian Z. Isolation and identification of yeast flora from genital tract in healthy female camels (*Camelus dromedarius*). *Veterinary Microbiology*. (2010); 144:183-186.

Shukla MK. *Applied Veterinary Andrology and Frozen Semen Technology*. New India Publishing Agency. (2011).

Thibier M and Guerin B. Hygienic aspects of storage and use of semen for artificial insemination. *Animal Reproduction Science*. (2000); 62:233-251.

Tibary A, Fite C, Anouassi A and Sghiri A. Infectious causes of reproductive loss in camelids. *Theriogenology*. (2006); 66(3):283-298.

Tingari MD, Manna MM, Rahim AT, Ahmed AK and Hamad MH. Studies on camel semen. I. Electroejaculation and some aspects of semen characteristics. *Animal Reproduction Science*. (1986); 12:213-222.

Waheed MM, Ghoneim IM and Alhaider AK. Seminal plasma and serum fertility biomarkers in dromedary camels (*Camelus dromedarius*). *Theriogenology*. (2015); 83:650-654.

Wernery U and Wernery R. Uterine infections in the dromedary camel - a review. *Proceedings of the First International Camel Conference, Dubai, UAE*. (1992); pp 155-158.

White PF, Way WL and Trevor AJ. Ketamine - Its pharmacology and therapeutic use. *Anaesthesiology*. (1982); 56:119-136.

Wickware CL, Johnson TA and Koziol JH. Composition and diversity of the preputial microbiota in healthy bulls. *Theriogenology*. (2020); 145:231-237.

Zhao XX. Reproduction in the Bactrian camel. In: Gahlot T.K. (editor), *Selected topics on camelids*. The camelid Publishers, Camel House 67, Gandhi Nagar West, Near Lalgarh Palace, Bikaner-334001, India, P, No. (2000); 499-538.